

Consideration of the ischaemic basis and treatment of Alzheimer's disease

Dedicated in Honor of my Mother Antonina Pluta (1922-2009) and Assistant Professor Irmina Zelman my Mentor and Friend (1927-2010)

Ryszard Pluta¹, Marzena Ułamek¹, Mirosław Jabłoński²

¹Laboratory of Ischemic and Neurodegenerative Brain Research, Department of Neurodegenerative Disorders, Mossakowski Medical Research Centre, Polish Academy of Sciences, Warsaw, Poland, ²Department of Orthopedics and Rehabilitation, Lublin Medical University, Lublin, Poland

Folia Neuropathol 2010; 48 (1): 11-26

Abstract

Victims of Alzheimer's disease (AD) develop a progressive dementia over years, accompanied by development of neurofibrillary tangles and finally neuronal death, accumulation of amyloid plaques and deposition of amyloid in neurovessels. Currently AD is the major form of dementia and the fourth leading cause of death in aged population. The investigation of etiology and therapy of AD, now more than ever, needs an infusion of new concepts. The aims of this review are to analyze knowledge of the influence of ischaemic and amyloid pathology on the final development of AD, especially with regards to the etiology of AD plaques, to develop a consensus on whether ischaemic blood-brain barrier permeability for amyloid peptide or both are a valid target for AD therapy. Reviewing experimental models of AD, we will address the issue whether plaques of amyloid persist, develop with time or both in animals during different forms of experimental therapy. Based on above suggestions recent direct evidence that amyloid plaques and neurofibrillary tangles can be cleared from the brain is thus provided in experimental condition. Moreover, recent study provides data that immunization with β -amyloid peptide decreases blood-brain barrier permeability for β -amyloid peptide decreases blood-brain barrier permeability for β -amyloid peptide or integrity. This review summarizes the latest advances in this area focusing on investigations based on in vivo animal studies.

Key words: Alzheimer's disease, ischaemic etiology, amyloid therapy, tau protein therapy, blood-brain barrier therapy, inflammation therapy, gene therapy, immunotherapy.

Communicating author:

Prof. Ryszard Pluta, MD, PhD, Laboratory of Ischemic and Neurodegenerative Brain Research, Department of Neurodegenerative Disorders, Mossakowski Medical Research Centre, Polish Academy of Sciences, Pawińskiego 5 Str., 02-106 Warsaw, Poland, phone +(48) (22) 60-86-540 or 60-86-469, fax +(48) (22) 668-55-32; e-mail: pluta@cmdik.pan.pl

Introduction

Around 6.2 million people in Europe are estimated to suffer from dementia of which Alzheimer's disease (AD) accounts for around three quarters of the cases. Taking into account their careers and families for whom caring often becomes a heavy personal and financial burden, some 20 million people are affected, i.e. around 4% of the European population. The number of patients with AD doubles every 5 years beyond age 65. Alzheimer's disease affects more than 25% of the age 85 and 40% of those aged 90 and over. Alzheimer's disease is the most common of dementias, accounting for over 60% of all cases over age 65. Now the population of Poland aged ≥65 years is 6.5 million. It is estimated that about 260 thousands individuals suffer from Alzheimer dementia in our country. The second most common form of dementias is vascular dementia, usually resulting from vascular brain diseases. Late onset AD is not caused by ageing, nor is it an inevitable part of the ageing process, it is age-related [6,169]. Alzheimer's disease affects circa 19 million people worldwide without developing countries with a prevalence of approximately 1 percent in the total population, although the risk of being afflicted with AD increases with age [6,132,169]. By 2025 the population of world aged 65 years and older will exceed one billion with more than 700 million living in developing countries. Now when the lifespan quickly increases, the number of sporadic AD cases increases dramatically, too. Actually almost 30 million individuals in around the world suffer from Alzheimer's dementia and this number of sick persons in the next century will multiply several fold unless treatments to prevent, or cure the disease that currently are unavailable will be found. Alzheimer's disease is already responsible for a huge social and economic costs projected to rise exponentially in the coming decades [157] as the elderly part of the society continues to increase [27].

Now it is clear that AD is multifactor [50,72,76, 119,168] and thus heterogeneous disease [50,71, 81,105]. Alzheimer's disease can affect different people in different ways but the most common symptom pattern begins with gradually worsening difficulty in remembering new information. This is because disruption of brain cells network, which usually begins in areas involved in the forming new memories. As the damage spreads, patients also experience confusion, disorganized thinking, impaired judgment, trouble

expressing themselves and disorientation with regard to location, time and space that may lead to unsafe wandering and socially inappropriate behavior. In advanced AD, patients need help bathing, dressing, using the bathroom, eating and carrying out other daily activities. Those in the final stages of the disease, lose their ability to communicate, fail to recognize loved ones and become bedbound and reliant on continual care. Alzheimer's disease is finally fatal. The course the disease takes and how fast changes occur vary from case to case. On average, sporadic AD patients live almost 10 years after they are diagnosed, though the disease can last for as many as 20 years [167].

Despite ongoing experimental and clinical interest in AD, the cause of the disease in sporadic cases is not known [7,67,109,144]. It is postulated that more than one pathogenetic pathway is involved in AD etiology. Owing to the involvement of more than one trigger factor in development Alzheimer's pathology it is important for the discovery specific therapies to prevent and treat this disease. Development of therapies requires the ability to correctly diagnose disease and knowledge about its neuropatogenesis. Diagnosis in AD patients is always late. The study of etiology and therapy [15] of sporadic AD, now more than ever needs an infusion of a new proposals. In 2007 approximately US\$ 5.5 billion was spent on the symptomatic treatment of AD. The vast majority of this expense was generated by just four drugs within two main classes, the acetylcholinesterase inhibitors [53] and N-methyl-D-aspartate receptor antagonists [148,165, 173]. Since all currently available treatments are symptomatic treatments, aimed at alleviating the symptoms of the disease and trying to slow the deterioration of the patients, there is a significant current need for improved drugs that can modify the underlying course of the disease. New concepts are needed as current obligatory treatment directed at symptomatic relief in AD patients has shown to be marginally effective or even a lack of efficacy has been demonstrated [2,15,53,116,148,173,178,179]. Despite of several expensive drugs worldwide for therapy of AD, the disease still robs millions aged individuals of both their memory and their live. New science is identifying many of the novel pathways [13,129,135,136,166, 168,169], which contribute to this damaging live disease, providing unprecedented opportunity for the discovery new therapies aimed at the root causes of AD [82,83,94,103]. More effective therapies directed at the cause of disease are needed. The genomic and ischaemic basis of AD will be defined completely in the near future, and corresponding molecular therapy targets will be identified. Ischaemic and genomic theories in brain degeneration have arrived and their application to resolving AD is our best hope.

The future front runner is autoimmunization, although this too is facing challenges in development. It may be some time before the first disease modifying agents emerge, which could revolutionize the way AD is treated. In 1998 [123], scientific interest in a new treatment approach to the therapy of AD was ignited after ischaemic model of AD investigation [120,122, 125], which indicated that it might be possible to immunize against the damaging properties of human β -amyloid peptide 1-42, [123,124] which led to accumulation and aggregation of this danger peptide in intra- and extracellular space of brain tissue [120,121, 124,126]. Pluta and colleagues [123] first presented that active immunization with human β -amyloid peptide 1-42 completely removed full-length amyloid from ischaemic rat brain [123,124]. This finding next has been partially confirmed and extended in other models of AD using both active [154] and passive [10] immunization. The main aim of this review is on the novel treatments for AD with a special emphasis on delivering against Alzheimer's proteins strategies. The expected epidemic number of AD cases in the next century makes the progress and discovery of effective therapy a matter of greatest importance and urgency. Presented review is good reason for revolutionary changes in future therapy of AD.

Ischaemic hypothesis of Alzheimer's disease

Recent study on transgenic AD animals with overexpression of amyloid precursor protein presented that cerebral blood flow is impaired in the animal models even before development of amyloid plaques and/or vascular amyloid deposition [69]. At least one third of brains with Alzheimer type dementia exhibit different neurovascular disorders [77]. In brains of AD cases, micro or macro intracerebral infarctions and white matter ischaemic damage [131,139] are evident [77]. Approximately 40% of cases with vascular type dementia had Alzheimer's-type pathology such as different kind of plaques [74,182], neurofibrillary tangles [80], hemorrhages [31,118] and neuronal death in hippocampus [75,76]. The presence of ischaemic changes seems usually ignored and regarded by scientists as insignificant or considered incidental in AD neuropathology. Interestingly, that Alzheimer in his original case of AD had made a note that besides "storage of peculiar material in the cortex, one sees endothelial proliferation and also occasionally neovascularisation" [3]. Endothelial proliferation with angiogenesis in the brain vessels of first patient provides data that ischaemic pathology was also present in the first case of AD. Above data suggest that we have overlap between vascular dementia and Alzheimer dementia [75,132] and that cerebrovascular pathology [13] plays a main role in the pathogenesis of AD. Aforementioned data raised the question what was the first: neurovascular disorder as a starter of AD [132] or degeneration of Alzheimer'stype itself [40]? Recent data propose a triggering and significant role for ischaemic mechanisms contributing to the degenerative processes in AD [13,77,113,125, 129,132,135,137,141,180,193]. Collected findings suggest that neuronal death following ischaemia with amyloid peptide from ischaemic circulatory system modulate ischaemic brain injury via molecular events in common with Alzheimer-type pathology [137 see for references]. These data indicate that brain ischaemia might be a key factor in the formation the full picture of Alzheimer dementia over years.

The brain has a limited response to different pathogens. For example similar neuropathological features are noted in the brain with ischaemia and that of AD. The pathogenesis of and a relationship between ischaemic dementia and Alzheimer's dementia are lastly much debated [40,76,113,119,128,137]. The role of both ischaemic brain injury and ischaemic blood-brain barrier changes in the pathology of AD is now more important than has long been assumed [19,22,113,119,125-130,133,135,136,163,191]. It is currently accepted that vascular dementia and AD share the same risk factors [132 see for references]. Cellular processes that lead to neurons demise in both disorders are known and are shared, too. Recently increasing information is mounting that pathological features of both disorders often occur concomitantly in individual cases. In fact mixed dementia may not represent two single co-occurring disorders but rather a single disease in which ischaemic hallmarks in neurons interact with focal ischaemic amyloid precursor protein metabolic alterations in characteristics brain regions. In contrast to the classical hypothesis of AD [6] new results indicate directly that brain ischaemia contributes to the progression of AD pathology [177]. Ischaemia is well known factor of neurons death, blood-brain barrier abnormalities, inflammatory response, tangles and plaques formation and finally dementia development [5,7,13,14,19,22,33,35,38,40, 50,51,58,68,74,76,78,80,86,113,137,161,162,177,180, 182,187,191,192]. Importantly aforementioned results showed those lesions, which mimic the biochemical and neuropathological changes as you can see in AD and they induce tau protein [180,187] and amyloid peptide pathologies and slow progressive cognitive impairment development [187].

Post mortem AD brain examination confirmed commonly present cerebrovascular alterations [5,185] that suggested ischaemic pathology. Epidemiological studies have presented a synergistic and spatial connection between neurovascular pathology and AD pathology [5] in formation the clinical evidences of dementia. For example nuns, who had neurovascular pathology, were more demented than those with tau pathology and a big number of amyloid plaques but without neurovascular pathology [161,162]. Additionally human Rotterdam investigation presented that clinically silent ischaemic brain injuries doubled the risk of dementia and had a direct connection with rapid cognitive decline as compared to individuals without ischaemic episodes [177]. Above data are supporting of the hypothesis that silent ischaemic injuries contribute to the Alzheimer phenotype dementia [132,177]. In summary, many AD cases had silent ischaemic episodes [177], which represent cause of neurons death, blood-brain barrier changes, inflammation response, neurofibrillary tangles and amyloid plaques formation and finally full-blown Alzheimer's dementia [5,7,13,14,19,22,33,35,38,40,50,51,58,68,74, 76,78,80,86,113,137,161,162,177,180,182,187,191,192].

Pathogenesis for AD neurodegeneration include: changes in calcium concentration [184], triggering of specific receptors affecting cell homeostasis, activation of oxidative processes [47], disruption of membrane integrity [96,175,176] and changes in lipid, influence of glutamate connected neurons death, inflammatory response, tau pathology [6,29,109] and pathological amyloid precursor protein processing [6,29,55,61,88,96,104,109,111,159] and aging [30,32, 98,155] or a combination of two or more of above processes. All above presented mechanisms you can observe in brain ischaemia. It has been suggested that ischaemic brain injury and ischaemic blood-brain barrier changes may have an important role in formation

brain degeneration with severe dementia [136 see for references]. It is well known that neurons death is occurring in brain ischaemia according to necrosis and apoptosis in brain sectors characteristic for AD such as entorhinal cortex and hippocampus [125,142]. These areas of the brain are involved in memory, thought and language. Most signaling factors, which trigger neurons death are known in ischaemia and are recognized as a ubiquitous signaling network, which links specific cell-surface receptors with the cell nucleus. In this rival theory, ischaemia of the brain is responsible for neurons death and ischaemic changes of blood-brain barrier for amyloid peptide movement from circulatory system into the brain and finally amyloid plaques formation [133,134,136,140 see for references]. It means that cross talk between ischaemic neuronal death and ischaemic blood-brain barrier injury exists in AD brain and could have significant implications for the triggering and maturation of AD [129,132,140-142]. Moreover, AD decreased length of capillaries in hippocampus correlates well with clinical pathology of Alzheimer dementia [9]. Oligomeric form of amyloid peptide through ischaemic neuronal toxicity and ischaemic vasculotoxic properties may provide an important link between AD and ischaemic neurodegeneration processes of this type of dementia [38,39,69,119-121,193]. Amyloid influence on blood-brain barrier vessels [1] is therefore one possible mechanism of blood-brain barrier damage [4] in AD [19,51,52,97,99,163]. Some studies indicate that 90% of human's amyloid plaques [80] and 80% of amyloid plagues in the transgenic AD models [43] are in direct contact with blood-brain barrier neurovessels [31,141,142] and they have spatial correlation with circulation network [5]. The reasons for AD are not known, but recent evidence strongly suggests that neurovascular ischaemic alterations play an important role in development this kind of disorder [12,14,38,40,68,69,76,78,86,113,122,125,128,129,132,13 3,135]. Some data from experimental models indicates that neurovascular insufficiency may be a trigger in AD and could provide link between this disorder and ischaemic brain injury [14,69,125,132,133]. β -amyloid peptide 1-42 influences the production of occludin and claudin tight junction proteins potentially affecting blood-brain barrier permeability [99]. In transgenic AD model with amyloid precursor overexpression amyloid positive microvessels showed endothelial cells apoptosis. Interesting findings in transgenic models of AD suggested that this disorder had not only a neurodegenerative but also neurovascular elements [69]. Brain hypoperfusion and changes in blood-brain barrier transport could impair amyloid peptide clearance [12] and lead to increased level of both soluble and oligomeric amyloid peptides in brain tissue [69,127,130,131,142]. In addition brain hypoperfusion can trigger ischaemic injury, which may act synergistically with AD elements [69,131,138] to exacerbate slowly a cognitive deficit. Alzheimer's disorder is associated with severe neuropathology in vessels [9,77,191,193], changes in function [69] and functional MRI [156] that suggest that changes in the cerebral blood flow may be a predictor for AD development [91]. Recently it is important to look for new, rival hypothesis such as ischaemic theory, which designs new strategies for AD etiology and finally treatment [14,40,68,69,76,113,119,132,138]. Ischaemic processes in neuropathogenesis of AD may have significant implications for therapy of neurons loss in this disorder. In this rival hypothesis neurotoxicity of amyloid peptide [6] will contribute partially, if at all to Alzheimer's disease brain degeneration.

In summary the start of AD pathology involves an initial neurons changes triggered by ischaemia [69,74,182], which leading to enhanced neurons vulnerability to β -amyloid peptide [84] and the ischaemic changes of the blood-brain barrier [97,120] vessels with leakage of serum borne amyloid peptide [120,160] into the brain parenchyma, activation of amyloid peptide dependent neurons injuries [84] and finally culminating in the development of different amyloid plaques [74,120,182] and end in full-blown AD [132,141,177,180,187] (Fig. 1). It is proposed that AD may be caused by silent ischaemic episodes [177] that attack and slowly steal the minds of its victims. Moreover, ischaemia increases the toxicity of amyloid peptide. Next possibility is that ischaemia increases the vulnerability of primarily ischaemic neuronal cells to $\beta\text{-amyloid}$ peptide neurotoxicity or accumulation of β -amyloid peptide increased ischaemic vulnerability [84]. According recent data the brain ischaemia age's brain 3.6 years each hour without treatment [153] and that explain age-dependent progression of Alzheimer's disease.

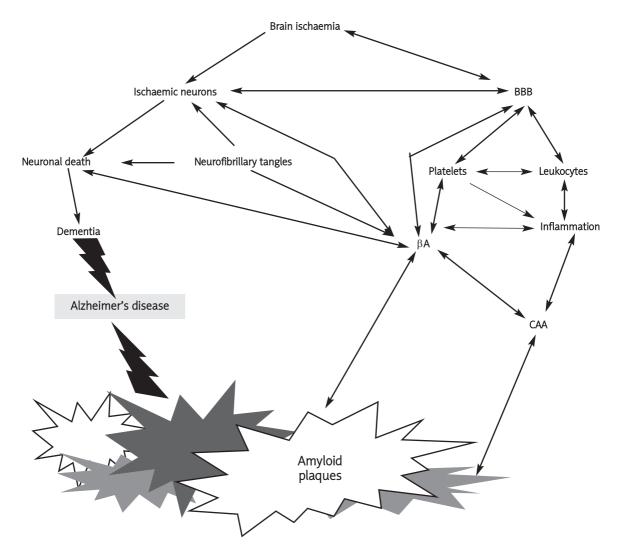
Anti-amyloid treatment

1998 is a turning point in the new history of novel idea in AD therapy [123]. At first the full success against human β -amyloid peptide 1-42 immunization

in ischaemic rats [123,124] and second moderate effect in transgenic mouse model overexpressing amyloid precursor protein [154] and third passive immunization against β -amyloid peptide [10] led to the fast development of immunotherapies in animals and clinical trials in human AD. In the past decade investigations on immunization had led to the formation of new experimental proposals as well as alternative routs of vaccine delivery to amyloid plaques in AD brain [27,62,64,82,83,94,103,164,165,172].

Human β -amyloid peptide clearance therapy had remarkable effects in ischaemic AD model [123,124] and less effect in transgenic AD model [154]. In human clinic, vaccination results were less conclusive [62,94,112]. Trials in patients with AD were prematurely stopped when 6% of vaccinated cases developed aseptic meningoencephalitis [54,112,117]. In addition only 20% of cases produced antibody against amyloid peptide [54]. Brains treated with vaccine presented post mortem less plaques in different brain regions, as well as presence of T lymphocytes. Moreover, antibody responders presented any improvement in clinical memory tests. Above results showed that vaccine therapy against β -amyloid peptide might still be a viable option for the treatment of individuals with AD. Currently, it has been observed that antibodies against β -amyloid peptide are present in human immunoglobulin that specifically recognize and inhibit the neurotoxic effects of β -amyloid peptide [44].

Over the past ten years ideas of therapy have been focused on inhibitors of β - and γ -secretases responsible for the production β -amyloid peptide from parent amyloid precursor protein [45,149,158]. Decrease of different forms of β -amyloid peptide in the brain of old rats after oral delivery of the γ -secretase inhibitors has been shown to reduce levels of β -amyloid peptides in both brain parenchyma and cerebrospinal fluid [17,49]. Another new line of investigation is the use of antibody against β -secretase in which reduction of different amyloid forms was observed in transgenic animals [145]. This reduction correlated with some improvement of cognitive activity. Two single-chain antibodies have been presented to possess α -secretase activity providing a novel use of immunotherapy [103,147]. Other scientists have used small particle libraries to screen for molecules, which either interfere with assembly of β -amyloid peptide particles into fibrils [37,89] or disaggregate existing fibrils [18,57,164].



 $\beta A - \beta$ -amyloid peptide, BBB – blood-brain barrier, CAA – cerebral amyloid angiopathy

Fig. 1. Ischaemic basis for sporadic Alzheimer's disease neuropathogenesis.

Neprilysin gene transfer into the brain leads to a remarkable reduction of β -amyloid peptide deposits in transgenic AD model [79,100]. Above data suggest that the deficient degradation of β -amyloid peptide caused by a low levels of neprilysin might contribute to the AD development. Insulin degrading enzyme is another major enzyme for β -amyloid peptide degradation in the brain [103]. Overexpression of this enzyme decreases level of β -amyloid peptide and retards or completely prevents amyloid plaques formation in the brain tissue [92]. Another two enzymes, angiotensin converting enzyme and endothelin converting enzyme degrade β -amyloid peptide, too [47,63]. Therapy by gelsolin, an agent that has a high affinity for β -amyloid peptide decreased the level of β -amyloid peptide in the brain tissue probably *via* a peripheral action [102]. Other β -amyloid peptide bindable drug, curcumin can cross blood-brain barrier and bind amyloid plaques and decrease amyloid concentration and plaque burden in transgenic model of AD [186]. The enoxaparin β -amyloid peptide bindable agent delivered peripherally significantly lowered β -amyloid peptide deposits in cortex and the total amyloid peptide cortical level probably by sequestering the serum β -amyloid peptide peripherally [16]. According to the peripheral sink theory [41,42], β -amyloid peptide bindable substances sequester serum β -amyloid peptide that leads to clearance of amyloid peptide by promoting a net efflux of a rapidly mobilized soluble pool of β -amyloid peptide. The peripheral sink theory was proposed by DeMattos *et al.*, [41] and was based on results obtained by passive immunization transgenic mice. These investigators proposed a model where sequestering of β -amyloid peptide in immune complexes in the plasma decreases the level of soluble β -amyloid peptide, which then contributes to a net efflux of amyloid peptide from the brain into blood [41,42].

Currently endogenous autoantibodies against β -amyloid peptide and receptor for advanced glycation-end-products peptides have been noted in healthy persons and individuals with AD [108]. Above data suggests that physiologically occurring autoantibodies against amyloid peptide and receptor for advanced glycation-end-products might be effective to β -amyloid peptide clearance from brain and blood.

Anti-tau treatment

Experimental treatments have been directed against hyperphosphorylated tau protein either by inhibiting various protein kinases or promoting phosphatase activities [70,83,90]. Current *in vitro* investigation shown small particles, which inhibited tau protein filament nucleation and fibrillization, making this substance a promising candidate to test in animal models of AD [25]. A new interesting observation about amyloid peptide vaccination has been presented lastly in experiments in which triple transgenic mice were passively immunized with antibodies against β -amyloid peptide [115]. β -amyloid peptide immunization leads to clearance of early but not late hyperphosphorylated tau protein aggregates *via* the proteasome [115].

Some investigation presented that memantine reversed hyperphosphorylation of tau protein in hippocampal slices [95] and this effect of memantine occurred by disinhibition of the function of protein phosphatase 2A [26] that earlier was noted to be downregulated in brains of AD patients [56]. Based on above information's, it was presented in human clinic that therapy of AD cases by memantine [148,173] during one year significantly decreases hyperphosphorylated tau in cerebrospinal fluid [59].

Anti-inflammatory treatment

In AD brains, the microglial cells behaved as inflammatory invaders, which cause an unintended pathology via release of cytokines designed to answer to primary brain pathology. This reaction may lead to a progression of AD through neurons death. Epidemiological observations suggest that long use of nonsteroidal anti-inflammatory therapy may prevent AD development [106,170]. Based on the above data, studies were undertaken to investigate the effects of anti-inflammation treatment in AD models [47]. These investigations include nonsteroidal anti-inflammatory drugs [107], peroxisome proliferator-activated receptor- γ agonists [46,65,152] and cannabinoids [146]. Another new data from transgenic model of AD showed that immunotherapy against β -secretase decreases development of inflammation in brain injury [145].

Umbilical cord blood cells delivered *i.v.* 48 hours after stroke are able to decrease neurodegeneration by providing neuroprotection and blocking the inflammatory reaction [181]. Injected cells appear to do this via multiple mechanisms: providing neuroprotection, modulating the inflammatory response, interrupting the apoptotic cascade, and enhancing neurogenesis and angiogenesis. Activation of sigma-1 and sigma-2 receptors via 1,3-di-0-tolylguanidin delivery 24 hours after stroke is equally impressive in reducing stroke injury [181]. Above molecule is neuroprotective by decreasing intracellular calcium in neuronal cells and inflammatory response by blocking the production of cytokines from brain immune cells. In brain the ischaemia and AD signals from the degenerating neuronal cells trigger immune cells for an inflammatory response, with increased production of cytokines. Whether the cause is known or not, all neurological diseases show similar intracellular neuronal signaling and inflammation reactions. The aforementioned therapeutic approaches may not only be beneficial for therapy of ischaemic stroke [181] but also for AD. Aforementioned two therapies act in a similar manner by inhibiting the peripheral immune system and promoting neuronal survival [181].

Blood-brain barrier treatment

The morphological and functional integrity of the brain depends on the coupling between cerebral blood flow and transport *via* the blood-brain barrier and neuronal function. In literature there are data

that neurovascular unit insufficiency may be triggered sporadic AD development [14,113,125,132, 137,163,193]. Possible etiological role of ischaemia in the development of AD have been presented in detail by several investigators [40,69,76,113,125,132,137]. Brain blood flow maintains a control of the neuronal environment not only by autoregulation of local blood flow but also by influencing transport by blood-brain barrier. The blood-brain barrier is a highly energetic system with different forms of transport via both its blood- and brain-facing sites. Structure of the blood facing side allows entry of nutrients products but opposite brain facing eliminate toxic products such as β -amyloid peptide from brain [34,35,126,193]. Among others an important role of the blood-brain barrier is control of the brain pool of β -amyloid peptide. The aim of this section of review is to analyze knowledge of the connection of the ischaemic blood-brain barrier with final formation AD, especially with regards to the development of amyloid plaques [132,136,137] and to develop a consensus on whether blood-brain barrier alterations are a valid target for AD treatment [135-138] and to stimulate scientists' discussion on the most important part of rival theory with regards to maturation of AD [132,133]. According to the ischaemic blood-brain barrier maturation theory of AD [133] all elements of blood-brain barrier such as endothelium, basal lamina, pericytes and astrocytes are main targets for therapy of AD [163]. The current idea states that injured blood-brain barrier transport system by ischaemia at its luminal and abluminal sides for β -amyloid peptide with damaged neurons by ischaemia are responsible for full-blown sporadic AD [128,129,133,135,136,140-142]. In this way new and more effective therapy approaches can be developed and more data on different amyloidosis can be gathered. Above observations suggest that stopping leakage of β -amyloid peptide from blood to brain tissue [43] and increasing reverse transport from brain into blood [12,126] and preventing ischaemic processes in neurons [137 see for references] are principal main points in treatment AD [20,24,33-36,60,73,78,85,106, 138,172]. Current studies provide data that active immunization with β -amyloid peptide reduces bloodbrain barrier permeability, amyloid burden and neuroinflammation as microgliosis in transgenic model of AD [43]. It was proved that the integrity of the bloodbrain barrier is disrupted in AD models and following β -amyloid peptide immunization the immune system clears amyloid peptide from sources in the brain tis-

sue as it would in peripheral organs lacking barriers. Once β -amyloid peptide is removed, the integrity of the blood-brain barrier is restored [43]. Above investigation clearly proves that the blood-brain barrier is disrupted in AD brain [22,163,191] and that immunization with β -amyloid peptide repairs the damage blood-brain barrier in transgenic AD model [43]. Earlier we have proved that active immunization with human β -amyloid peptide 1-42 in ischaemic model of AD reverses the ischaemic blood-brain barrier permeability for β -amyloid peptide 1-42 [126] and prevent further disease progression [123,124]. One possible explanation of the restoration of the blood-brain barrier is that the active immunization leads to the reduction in the level of circulating β -amyloid peptide [43], which could directly or indirectly influence the activity of the blood-brain barrier [12,51,99]. For example, inflammatory factors such as IL-1 β , IL-6 and TNF- α [21,151] that stimulate angiogenesis [58] and β -amyloid peptide have been noticed to influence an increase of some angiogenic factors like TGF- β and VEGF [143,174]. It can be concluded that with the removal of informatory substances provided by β -amyloid peptide, the endothelium become intact and tight junctions closed, thereby restoring a physiological blood-brain barrier activity. Increased level of β -amyloid peptide in plasma has been observed in a transgenic model of AD after active immunization with amyloid and *i.v.* delivery of molecules with an affinity to β -amyloid peptide [42,102] and after active immunization [123,124] of non-human primates [93]. It is proposed that molecules that sequester serum β -amyloid peptide may decrease or prevent brain amyloidosis [102]. Finally investigations with antibodies against intercellular adhesion molecule-1 [189] or platelet-endothelial cell adhesion molecule-1 [150] have shown that blockage of adhesion molecules and/or leukocyte adhesion [28] or platelets (> 90% of β -amyloid peptide in blood is stored in platelets) attachment respectively reduces brain neurodegeneration after effects.

Several ways have been proposed to clear out β -amyloid peptide through blood-brain barrier including: especially receptor-mediated β -amyloid peptide reverse transport *via* blood-brain barrier [126], enzyme mediated β -amyloid peptide degradation and β -amyloid peptide bindable molecule mediated β -amyloid peptide clearance. Receptor mediated transport of β -amyloid peptide *via* blood-brain barrier is mainly responsible for both efflux and influx of amyloid peptide. Lipoprotein receptor-related protein mediates efflux of β -amyloid peptide from brain into blood [12,35]. The interaction between lipoprotein receptor-related protein and amyloid mediates β -amyloid peptide blood-brain barrier vessels binding, endocytosis and transcytosis through blood-brain barrier into circulatory system [66]. Additionally p-glycoprotein has been proposed to be involved in amyloid clearance via blood-brain barrier [87]. Currently, some results suggest that the neonatal Fc receptor at the blood-brain barrier plays a crucial role in IgGassisted β -amyloid removal from the aging brain [36]. Receptor for advanced glycation-end-products mediates influx of amyloid from serum into brain tissue [33,35]. Downregulation of receptor for advanced glycation-end-products can inhibit influx of amyloid peptide [33]. Gp 330/megalin has been noted to transport blood β -amyloid peptide in a complex with apolipoprotein J into brain tissue through blood-brain barrier [192]. Lipoprotein receptor-related protein and receptor for advanced glycation-end-products play opposing roles in β -amyloid peptide transport through blood-brain barrier [35]. One important strategy would be to discover new drugs, which regulate the function or expression of amyloid transport receptors via blood-brain barrier vessels. The decreased regulation of receptor for advanced glycation-endproducts and increased regulation of lipoprotein receptor-related protein in blood-brain barrier might readjust the transport equilibrium for amyloid by promoting its net efflux from brain into plasma. Statins, which increased lipoprotein receptor-related protein in blood-brain barrier, might facilitate the clearance of amyloid from brain tissue [34]. It is worth noting that receptor for advanced glycation-end-products blockades using receptor for advanced glycation-endproducts specific IgG [108] can also increase the expression of lipoprotein receptor-related protein [34].

Gene treatment

Gene treatments are completely new forms of therapy in which genes are transferred into the damaged cells [64]. Current results suggest that damaged genes in ischaemic brain injury might be harmful to different kinds of brain cells. Thus gene treatment may serve to rescue those cells from potential cells death. In this respect, recent developments have presented medical effects of gene treatment in experimental global and focal ischaemic models [114]. Using gene therapy against apoptosis may reduce apoptotic cells following ischaemic brain injury [23,190]. Beneficial results were noticed by inhibiting apoptosis and enhancing glial cell survival following ischaemic neurodegeneration [183]. Moreover, midkine and heat shock protein gene transfer showed neuroprotection in ischaemic brain [8,171]. As such, gene treatment offers new interesting and powerful therapies in the future for ischaemic stroke and AD patients reducing amyloid plaque burden *via ex vivo* gene delivery of an amyloid degrading protease [64]. As example neprilysin gene transfer reduces human amyloid pathology in transgenic mice [79,100].

Other treatments

Scientific data suggest that ischaemic neuronal cells undergo necrosis and apoptosis, necroptosis and autophagic programmed cell death, which finally leads to neurodegeneration with dementia. The important factors in apoptosis are tumor necrosis factor-receptor-1 and CD95, which in ischaemic brain are overexpressed. Thus, influence on tumor necrosis factor-receptor-1 and CD95 by antibody treatment will induce neuroprotection in brain ischaemia. On the basis of this idea *i.v.* therapy with antibodies against tumor necrosis factor- α and/or CD95L significantly reduced the infarct volume in experimental brain ischaemia [11,110] and mortality [101]. These results suggest that blocking tumor necrosis factor and CD95L activity with antibodies, we can prevent primary and secondary responses to ischaemia and probably to AD.

Conclusions

Considerable progress has been made in the last decade by handful scientists in resolving the etiology and the development of new perspective therapies for AD. Efforts to intensification of therapy research are justified at both humanistic and economic levels [27,157]. Extensive investigation designed to discovery new ways to delay onset and progression of AD and on new therapies recently is ongoing worldwide [103]. Since all degenerative diseases share commonalities that lead to neurons death, these treatments proposals not only apply to ischaemic model of AD [120,122,125,132,138,141,142] but to AD transgenic models [94,166]. Scientists are evaluating other

potentially promising approaches studying e.g. the active and passive immunization treatments and trying combinations of therapies and investigating relationships between ischaemic and Alzheimer's dementia therapies. Among treatments for AD in both experimental and clinical trials are many strategies to block toxic β -amyloid peptide 1-42 and to rescue the vulnerable neuronal cells from death. Other proposals aim to prevent the co-pathogenic effects of different proteins e.g. amyloid and tau protein [6,83]. New insights into selective neurons vulnerability and the link between brain ischaemia and AD may provide novel entry points for effective therapy. By controlling blood-to-brain and brain-to-blood β -amyloid peptide movement [35,120,126] the blood-brain barrier may self-limit amyloid dependent capillary injuries and decrease the risk of neurons death and β -amyloid plaques formation [132,137,140]. It can be concluded that immunotherapy and other treatments based on β -amyloid peptide removal from brain may be beneficial in limiting the degree of secondary degeneration caused by amyloid properties [6]. Enhancement of β -amyloid peptide degradation enzymes via gene therapy, transcriptional activation or even pharmacological activation of the β -amyloid peptide degrading enzymes represents a new therapeutic proposal for the treatment of AD [48]. Based on the peripheral sink theory, it is possible to decrease brain β -amyloid peptide burden without the need for antibodies and therapeutic agents to move via the blood-brain barrier [41,42]. These findings indicated that treatment against β -amyloid peptide might still be a viable option for the treatment of AD, if potentially harmful proinflammatory effects can be avoided. These data also suggest that stopping leakage of β -amyloid peptide from blood into brain [43] and increasing its reverse movement from brain tissue into blood [12,126] can help individuals with AD [34,42,73,85, 172].

Recently it has been difficult in translating experimental treatments into effective clinical cure. The issue is that investigators do not consider important variables in dealing with human cases as opposed to rodent models. Actually the standard in developing drugs has been to focus on a single target. However, the underlying degenerative pathways in neurological disorders involve an intertwining of many mechanisms and treating one will not necessarily change the outcome of the disorder. The current focus is shifting to a multi-functional approach in which a single drug has multiple neurobiochemical targets and can therefore cure the disorder or consequences of pathology more fully [188]. In the future treatment proposals will likely address events, which are upstream of a more broadly construed pathological cascade, which includes but is not limited to the production and deposition of β -amyloid peptide.

Acknowledgements

This study was supported in part by founds from: Mossakowski Medical Research Centre (T5), Polish Ministry of Science and Higher Education (2007-2010-Cost/253/2006) and European Union (Cost Action B30).

References

- Alonzo NC, Hyman BT, Rebeck GW, Greenberg SM. Progression of cerebral amyloid angiopathy: accumulation of amyloid-beta 40 in affected vessels. J Neuropathol Exp Neurol 1998; 57: 353-359.
- Alvarez XA, Cacabelos R, Laredo M, Couceiro V, Sampedro C, Varela M, Corzo L, Fernandez-Novoa L, Vargas M, Aleixandre M, Linares C, Granizo E, Muresanu D, Moessler H. A 24-week, double-blind, placebo-controlled study of three dosages of Cerebrolysin in patients with mild to moderate Alzheimer's disease. Eur J Neurol 2006; 13: 43-54.
- Alzheimer A. Über eine eigenartige Erkrankung der Hirnrinde. Allg Z Psychiatr Psych Gericht Med (Berlin) 1907; 64: 146-148.
- 4. Anfuso CD, Assero G, Lupo G, Nicota A, Cannavo G, Strosznajder RP, Rapisarda P, Pluta R, Alberghina M. Amyloid beta (1-42) and its beta (25-35) fragment induce activation and membrane translocation of cytosolic phospholipase A(2) in bovine retina capillary pericytes. Biochim Biophys Acta 2004; 1686: 125-138.
- 5. Armstrong RA. Spatial correlation between β -amyloid (A β) deposits and blood vessels in familial Alzheimer's disease. Folia Neuropathol 2008; 46: 241-248.
- Armstrong RA. The molecular biology of senile plaques and neurofibrillary tangles in Alzheimer's disease. Folia Neuropathol 2009; 47: 289-299.
- 7. Ashe KH. Learning and memory in transgenic mice modeling Alzheimer's disease. Learn Mem 2001; 8: 301-308.
- Badin RA, Lythgoe MF, Van der Weerd L, Thomas DL, Gadian DG, Latchman DS. Neuroprotective effects of virally delivered HSPs in experimental stroke. J Cereb Blood Flow Metab 2006; 26: 371-381.
- 9. Bailey TL, Rivara CB, Rocher AB, Hof PR. The nature and effects of cortical microvascular pathology in aging and Alzheimer's disease. Neurol Res 2004; 26: 573-578.
- Bard F, Cannon C, Barbour R, Burke RL, Games D, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Lieberburg I, Motter R, Nguyen M, Soriano F, Vasquez N, Weiss K, Welch B, Seubert P, Schenk D, Yednock. Peripherally administered antibodies against amyloid beta-

peptide enter the central nervous system and reduce pathology in a mouse model of Alzheimer disease. Nat Med 2000; 6: 916-919.

- Barone FC, Arvin B, White RF, Miller A, Webb CL, Willette RN, Lysko PG, Feuerstein GZ. Tumor necrosis factor-alpha. A mediator of focal ischemic brain injury. Stroke 1997; 28: 1233-1244.
- 12. Bell RD, Deane R, Chow N, Long X, Sagare A, Singh I, Streb JW, Guo H, Rubio A, Van Nostrand W, Miano JM, Zlokovic BV. SRF and myocardin regulate LRP-mediated amyloid-beta clearance in brain vascular cells. Nat Cell Biol 2009; 11: 143-153.
- 13. Bell RD, Zlokovic BV. Neurovascular mechanisms and bloodbrain barrier disorder in Alzheimer's disease. Acta Neuropathol 2009;118: 103-113.
- 14. Benarroch E. Neurovascular unit dysfunction: A vascular component of Alzheimer disease? Neurology 2007; 68: 1730-1732.
- Bennett DA, Wilson RS, Schneider JA, Evans DA, Mendes de Leon CF, Arnold SE, Barness LL, Bienias JL. Education modifies the relation of AD pathology to level of cognitive function in older persons. Neurology 2003; 60: 1909-1915.
- 16. Bergamaschini L, Rossi E, Storini C, Pizzimenti S, Distaso M, Pergo C, De Luigi A, Vergani C, De Simoni MG. Peripheral treatment with enoxaparin, a low molecular weight heparin, reduces plaques and beta-amyloid accumulation in a mouse model of Alzheimer's disease J Neurosci 2004; 24: 4181-4186.
- 17. Best JD, Jay MT, Out F, Churcher I, Reilly M, Morentin-Gutierrez P, Pattison C, Harrison T, Shearman MS, Atack JR. In vivo characterization of A β (40) changes in brain and cerebrospinal fluid using the novel γ -secretase inhibitor N-[cis-4-[(4-chlorophenyl)sulfonyl]-4-(2,5-difluorophenyl)cyclohexyl]-1,1,1-trifluoromethane-sulfonamide (MRK-560) in the rat. J Pharmacol Exp Ther 2006; 317: 786-790.
- Blanchard BJ, Chen A, Rozenboom LM, Stafford KA, Weigele P, Ingram VM. Efficient reversal of Alzheimer's disease fibril formation and elimination of neurotoxicity by a small molecule. Proc Natl Acad Sci USA 2004; 101: 14326-14332.
- Blennow K, Wallin A, Fredman P, Karlsson I, Gottfries CG, Svennerholm L. Blood-brain barrier disturbance in patients with Alzheimer's disease is related to vascular factors. Acta Neurol Scand 1990; 81: 323-326.
- 20. Borlongan CV, Lind JG, Dillon-Carter O, Yu G, Hadman M, Cheng C, Carroll J, Hess DC. Bone marrow grafts restore cerebral blood flow and blood-brain barrier in stroke rats. Brain Res 2004; 1010: 108-116.
- 21. Boutin H, LeFeuvre RA, Horai R, Asano M, Iwakura Y, Rothwell NJ. Role of IL-1alpha and IL-1beta in ischemic brain damage. J Neurosci 2001; 21: 5528-5534.
- Bowman GL, Kaye JA, Moore M, Waichunas D, Carlson NE, Quinn JF. Blood-brain barrier impairment in Alzheimer disease: stability and functional significance. Neurology 2007; 68: 1809-1814.
- Cao YJ, Shibata T, Rainov NG. Liposome-mediated transfer of the bcl-2 gene results in neuroprotection after in vivo transient focal cerebral ischemia in an animal model. Gene Ther 2002; 9: 415-419.
- 24. Cheng T, Liu D, Griffin JH, Fernandez JA, Castellio F, Rosen ED, Fukudome K, Zlokovic BV. Activated protein C blocks p53-

mediated apoptosis in ischemic human brain endothelium and is neuroprotective. Nat Med 2003; 9: 338-342.

- 25. Chirita C, Necula M, Kuret J. Ligand-dependent inhibition and reversal of tau filament formation. Biochemistry 2004; 43: 2879-2887.
- 26. Chohan MO, Khatoon S, Grundke-Iqbal IG, Iqbal K. Involvement of I2PP2A in the abnormal hyperphosphorylation of tau and its reversal by memantine. FEBS Lett 2006; 580: 3973-3979.
- 27. Christensen DD. Changing the course of Alzheimer's disease: Anti-amyloid disease-modifying treatments on the horizon. Prim Care Companion J Clin Psychiatry 2007; 9: 32-41.
- 28. Clark WM, Madden KP, Rothlein R, Zivin J. Reduction of central nervous system ischemic injury in rabbits using leukocyte adhesion antibody treatment. Stroke 1991; 22: 877-883.
- 29. Clark CM, Xie S, Chittams J, Ewbank D, Peskind E, Galasko D, Morris JC, McKeel DWJr, Farlow M, Weitlauf SL, Quinn J, Kaye J, Knopman D, Arai H, Doody RS, DeCarli C, Leight S, Lee VH, Trojanowski JQ. Cerebrospinal fluid tau and β -amyloid: how well do these biomarkers reflect autopsy-confirmed dementia diagnoses? Arch Neurol 2003; 60: 1696-1702.
- 30. Corrada MM, Head E, Kim R, Kawas C. Braak and Braak staging and dementia in the oldest-old: preliminary results from the 90+ study. Neurology 2005; 64: A276.
- 31. Cullen KM, Kocsi Z, Stone J. Pericapillary haem-rich deposits evidence for microhaemorrhages in aging human cerebral cortex. J Cereb Blood Flow Metab 2005; 25: 1656-1667.
- Davis DG, Schmidt FA, Wekstein DR, Markesbery WR. Alzheimer neuropathologic alterations in aged cognitively normal subjects. J Neuropathol Exp Neurol 1999; 58: 376-388.
- 33. Deane R, Du Yan S, Submamaryan RK, LaRue B, Jovanovic S, Hogg E, Welch D, Manness L, Lin C, Yu J, Zhu H, Ghiso J, Frangione B, Stern A, Schmidt AM, Armstrong DL, Arnold B, Liliensiek B, Nawroth P, Hofman F, Kindy M, Stern D, Zlokovic BV. RAGE mediates amyloid-beta peptide transport across the blood-brain barrier and accumulation in brain. Nat Med 2003; 9: 907-913.
- 34. Deane R, Wu Z, Sagare A, Davis J, Du Yan S, Hamm K, Xu F, Parisi M, LaRue B, Hu HW, Spijkers P, Guo H, Song X, Lenting PJ, Van Nostrand WE, Zlokovic BV. LRP/amyloid beta-peptide interaction mediates differential brain efflux of Abeta isoforms. Neuron 2004; 43: 333-344.
- Deane R, Wu Z, Zlokovic BV. RAGE (yin) versus LRP (yang) balance regulates alzheimer amyloid beta-peptide clearance through transport across the blood-brain barrier. Stroke 2004; 35 (11 Suppl 1): 2628-2631.
- 36. Deane R, Sagare A, Hamm K, Parisi M, LaRue B, Guo H, Wu Z, Holtzman DM, Zlokovic BV. IgG-assisted age-dependent clearance of Alzheimer's amyloid beta peptide by the blood-brain barrier neonatal Fc receptor. J Neurosci 2005; 25: 11495-11503.
- 37. De Felice FG, Vieira MN, Saraiva LM, Figueroa-Villar JD, Garcia-Abreu J, Liu R, Chang L, Klein WL, Ferreira ST. Targeting the neurotoxic species in Alzheimer's disease: inhibitors of Abeta oligomerization. FASEB J 2004; 18: 1366-1372.
- 38. De la Torre JC. Vascular basis of Alzheimer's pathogenesis. Ann NY Acad Sci 2002; 977: 196-215.

- 39. De la Torre JC. Alzheimer's disease is a vasocognopathy: a new term to describe its nature. Neuronal Res 2004; 26: 517-524.
- 40. De la Torre JC. Is Alzheimer's disease preceded by neurodegeneration or cerebral hypoperfusion? Ann Neurol 2005; 57: 783-784.
- 41. DeMattos RB, Bales KR, Cummins DJ, Dodart JC, Paul SM, Holtzman DM. Peripheral anti-A beta antibody alters CNS and plasma A beta clearance and decrease brain A beta burden in a mouse model of Alzheimer's disease. Proc Natl Acad Sci USA 2001; 98: 8850-8855.
- 42. DeMattos RB, Bales KR, Cummins DJ, Paul SM, Holtzman DM. Brain to plasma amyloid- efflux: a measure of brain amyloid burden in a mouse model of Alzheimer's disease. Science 2002; 295: 2264-2267.
- Dickstein DL, Biron KE, Ujiie M, Pfeifer CG, Jeffries AR, Jefferies WA. Abeta peptide immunization restores blood-brain barrier integrity in Alzheimer disease. FASEB J 2006; 20: 426-433.
- 44. Dodel RC, Du Y, Depboylu C, Hampel H, Frolich L, Haag A, Hemmeter U, Paulsen S, Teipel SJ, Brettschneider S, Spottke A, Nolker C, Moller HJ, Wie X, Farlow M, Sommer N, Oertel WH. Intravenous immunoglobulins containing antibodies against beta-amyloid for the treatment of Alzheimer's disease. J Neurol Neurosurg Psychiatry 2004; 75: 1472-1474.
- 45. Dovey HF, John V, Anderson JP, Chen LZ, de Saint Andrieu P, Fang LY, Freedman SB, Folmer B, Goldbach E, Holsztynska EJ, Hu KL, Johnson-Wood KL, Kennedy SL, Kholodenko D, Knops JE, Latimer LH, Lee M, Liao Z, Lieberburg IM, Motter RN, Mutter LC, Nietz J, Quinn KP, Sacchi KL, Seubert PA, Shopp GM, Thorsett ED, Tung JS, Wu J, Yang S, Yin CT, Schenk DB, May PC, Altstiel LD, Bender MH, Boggs LN, Britton TC, Clemens JC, Czilli DL, Dieckman-McGinty DK, Droste JJ, Fuson KS, Gitter BD, Hyslop PA, Johnstone EM, Li WY, Little SP, Mabry TE, Miller FD, Audia JE. Functional gamma-secretase inhibitors reduce betaamyloid peptide levels in the brain. J Neurochem 2001; 76: 173-181.
- 46. Echeverria V, Clerman A, Dore S. Stimulation of PGE2 receptors EP2 and EP4 protects cultured neurons against oxidative stress and cell death following beta-amyloid exposure. Eur J Neurosci 2005; 22: 2199-2206.
- Eckman EA, Watson M, Marlow L, Sambamurti K, Eckman CB. Alzheimer's disease beta-amyloid peptide is increased in mice deficient in endothelin-converting enzyme. J Biol Chem 2003; 278: 2081-2084.
- Eckman EA, Eckman CB. Abeta-degrading enzymes: modulators of Alzheimer's disease pathogenesis and targets for therapeutic intervention. Biochem Soc Trans 2005; 33: 1101-1105.
- 49. El Mouedden M, Vandermeeren M, Meet T, Mercken M. Reduction of Abeta levels in the Sprague Dawley rat after oral administration of the functional gamma-secretase inhibitor, DAPT: a novel non-transgenic model for Abeta production inhibitors. Curr Pharm Des 2006; 12: 671-676.
- 50. Etiene D, Kraft J, Ganju N, Gomez-Isla T, Gemelli B, Hyman BT, Hedley-Whyte ET, Wands JR, De La Monte SM. Cerebrovascular pathology contributes to the heterogeneity of Alzheimer's disease. J Alzheimers Dis 1998; 1: 119-134.
- 51. Farkas IG, Czigner A, Farkas E, Dobo E, Soos K, Penke B, Endresz V, Mihaly A. Beta-amyloid peptide-induced blood-brain barrier

disruption facilitates T-cell entry into the rat brain. Acta Histochem 2003; 1005: 115-125.

- 52. Fiala M, Zhang L, Gan X, Sherry B, Taub D, Graves MC, Hama S, Way D, Weinand M, Witte M, Lorton D, Kuo YM, Roher AE. Amyloid-beta induces chemokine secretion and monocyte migration across a human blood-brain barrier model. Mol Med 1998; 4: 480-489.
- 53. Fillit HM, Doody Smith R, Binaso KB, Crooks GM, Ferris SH, Farlow MR, Leifer B, Mills Ch, Minkoff N, Orland B, Reichman WE, Salloway S. Recommendations for best practices in the treatment of Alzheimer's disease in managed care. Am J Geriatr Pharmacother 2006; 4 Suppl A: S9-S24.
- 54. Gilman S, Koller M, Black RS, Jenkins L, Griffith SG, Fox NC, Eisner L, Kirby L, Rovira MB, Forette F, Orgogozo JM, AN1792 (QS-21)-201 Study Team. Clinical effects of Abeta immunization (AN1792) in patients with Alzheimer's disease in an interrupted trial. Neurology 2005; 64: 1553-1562.
- 55. Glenner GG, Wong CW. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. Biochem Biophys Res Commun 1984; 120: 885-890.
- 56. Gong CX, Singh TJ, Grundke-Iqbal I, Iqbal K. Phosphoprotein phosphatase activities in Alzheimer disease. J Neurochem 1993; 61: 921-927.
- 57. Gong Y, Chang L, Viola KL, Loacor PN, Lambert MP, Finch CE, Krafft GA, Klein WL. Alzheimer's disease-affected brain: presence of oligomeric A beta ligands (ADDLs) suggests a molecular basis for reversible memory loss. Proc Natl Acad Sci USA 2003; 100: 10417-10422.
- Grammas P, Ovase R. Inflammatory factors are elevated in brain microvessels in Alzheimer's disease. Neurobiol Aging 2001; 22: 837-842.
- 59. Gunnarsson MD, Kilander L, Sudelof J, Basun H, Lannfelt L. Reduction of hyperphosphorylated tau during memantine treatment of Alzheimer's disease. Alzheimer's & Dementia Suppl. 2006; 2: S63-S64.
- 60. Guo H, Liu D, Gelbard H, Cheng T, Insalaco R, Fernandez JA, Griffin JH, Zlokovic BV. Activated protein C prevents neuronal apoptosis via protease-activated receptors 1 and 3. Neuron 2004; 41: 563-572.
- 61. Haroutunian V, Perl DP, Dushyant P, Purohit DP, Marin D, Khan K, Lantz M, Davis KL, Mohs RC. Regional distribution of neurotic plaques in the nondemented elderly and subjects with very mild Alzheimer's disease. Arch Neurol 1998; 55: 1185-1191.
- 62. Hawkes ChA, McLaurin J. Immunotherapy as treatment for Alzheimer's disease. Expert Rev Neurotherapeutics 2007; 7: 1535-1548.
- 63. Hemming ML, Selkoe DJ. Amyloid beta-protein is degraded by cellular angiotensin-converting enzyme (ACE) and elevated by an ACE inhibitor. J Biol Chem 2005; 280: 37644-37650.
- 64. Hemming ML, Patterson M, Reske-Nielsen C, Lin L, Isacson O, Selkoe DJ. Reducing amyloid plaque burden via ex vivo gene delivery of an Abeta-degrading protease: a novel therapeutic approach to Alzheimer disease. PLoS Med 2007; 4: 1405-1416.
- 65. Heneka MT, Sastre M, Dumitrescu-Ozimek L, Hanke A, Dewachter I, Kuiperi C, O'Banion K, Klockgether T, Van Leuven F, Landreth GE. Acute treatment with the PPARγ agonist piogli-

tazone and ibuprofen reduces glial inflammation and A β 1-42 levels in APPV7171 transgenic mice. Brain 2005; 128: 1442-1453.

- 66. Herz J. LRP: a bright beacon at the blood-brain barrier. J Clin Invest 2003; 112: 1483-1485.
- 67. Higgins GA, Jacobsen H. Transgenic mouse models of Alzheimer's disease: phenotype and application. Behav Pharmacol 2003; 14: 419-438.
- Humpel C, Marksteiner J. Cerebrovascular damage as a cause for Alzheimer's disease. Curr Neurovasc Res 2005; 2: 341-347.
- 69. ladecola C. Neurovascular regulation in the normal brain and in Alzheimer's disease. Nat Rev Neurosci 2004; 5: 347-360.
- Iqbal K, Grudke-Iqbal I. Inhibition of neurofibrillary degeneration: a promising approach to Alzheimer's disease and other tauopathies. Curr Drug Targets 2004; 5: 495-502.
- Iqbal K, Flory M, Khatoon S, Soininen H, Pirttila T, Lehtovirta M, Alafuzoff I, Blennow K, Andreasen N, Vanmechelen E, Grundke-Iqbal I. Subgroups of Alzheimer's disease based on cerebrospinal fluid molecular markers. Ann Neurol 2005; 58: 748-757.
- Iqbal K, Grundke-Iqbal I. Metabolic/signal transduction hypothesis of Alzheimer's disease and other tauopathies. Acta Neuropathol 2005; 109: 25-31.
- 73. Iwata N, Tsubuki S, Takaki Y, Shirotani K, Lu B, Gerard NP, Gerard C, Hama E, Lee HJ, Saido TC. Metabolic regulation of brain Abeta by neprilysin. Science 2001; 292: 1550-1552.
- 74. Jendroska K, Hoffmann OM, Patt S. Amyloid beta peptide and precursor protein (APP) in mild and severe brain ischemia. Ann NY Acad Sci 1997; 826: 401-404.
- 75. Kalaria RN, Ballard C. Overlap between pathology of Alzheimer's disease and vascular dementia. Alzheimer Dis Assoc Disord 1999; 13: S115-S123.
- 76. Kalaria RN. The role of cerebral ischemia in Alzheimer's disease. Neurobiol Aging 2000; 21: 321-330.
- 77. Kalaria RN. Small vessel disease and Alzheimer's dementia: Pathological considerations. Crebrovasc Dis 2002; 13 Suppl 2: 48-52.
- 78. Kalback W, Esh C, Castano EM, Rahman A, Kokjohn T, Luehrs DC, Sue L, Cisneros R, Gerber F, Richardson C, Bohrmann B, Walker DG, Beach TG, Roher AE. Atherosclerosis, vascular amyloidosis and brain hypoperfusion in the pathogenesis of sporadic Alzheimer's disease. Neurol Res 2004; 26: 525-539.
- 79. Kanemitsu H, Tomiyama T, Mori H. Human neprilysin is capable of degrading amyloid beta peptide not only in the monomeric form but also the pathological oligomeric form. Neurosci Lett 2003; 350: 113-116.
- Kawai M, Kalaria RN, Harik SI, Perry G. The relationship of amyloid plaques to cerebral capillaries in Alzheimer's disease. Am J Pathol 1990; 137: 1435-1446.
- 81. Katzman R, Terry R, DeTeresa R, Brown T, Davies P, Fuld P, Renbing X, Peck A. Clinical, pathological and neurochemical changes in dementia: a subgroup with preserved mental status and numerous neocortical plaques. Ann Neurol 1988; 23: 138-144.
- Kim HD, Jin JJ, Maxwell A, Fukuchi K. Enhancing Th2 immune responses against amyloid protein by a DNA prime-adenovirus boost regimen for Alzheimer's disease. Immunol Lett 2007; 112: 30-38.

- Klafki HW, Staufenbiel M, Kornhuber J, Wiltfang J. Therapeutic approaches to Alzheimer's disease. Brain 2006; 129: 2840-2855.
- 84. Koistinaho M, Kettunen MI, Goldsteins G, Keinanen R, Salminen A, Ort M, Bures J, Liu D, Kauppinen RA, Higgins LS, Koistinaho J. Beta-amyloid precursor protein transgenic mice that harbor diffuse A beta deposits but do not form plaques show increased ischemic vulnerability: role of inflammation. Proc Natl Acad Sci USA 2002; 99: 1610-1615.
- 85. Koistinaho M, Lin S, Wu X, Esterman M, Koger D, Hanson J, Higgs R, Liu F, Malkani S, Bales KR, Paul SM. Apolipoprotein E promotes astrocyte colocalization and degradation of deposited amyloid-beta peptides. Nat Med 2004; 10: 719-726.
- 86. Kudo T, Imaizumi K, Tanimukai H, Katayama T, Sato N, Nakamura Y, Tanaka T, Kashiwagi Y, Jinno Y, Tohyama M, Takeda M. Are cerebrovascular factors involved in Alzheimer's disease? Neurobiol Aging 2000; 21: 215-224.
- Lam FC, Liu R, Lu P, Shapiro AB, Renoir JM, Sharoni FJ, Reiner PB. Beta-amyloid efflux mediated by p-glycoprotein. J Neurochem 2001; 76: 1121-1128.
- Larson EB, Wang L. Research and practice in Alzheimer's disease. Vol. 12. Serdi Publisher, Paris 2007; pp. 38-41.
- Lashuel HA, Hartley DM, Balakhaneh D, Aggarwal A, Teichberg S, Callaway DJ. New class of inhibitors of amyloid-beta fibril formation. Implications for the mechanism of pathogenesis in Alzheimer's disease. J Biol Chem 2002; 277: 42881-42890.
- 90. Lau LF, Schachter JB, Seymour PA, Sanner MA. Tau protein phosphorylation as a therapeutic target in Alzheimer's disease. Curr Top Med Chem 2002; 2: 395-415.
- 91. Lee BC, Mintun M, Buckner RL, Morris JC. Imaging of Alzheimer's disease. J. Neuroimaging 2003; 13: 199-214.
- 92. Leissring MA, Farris W, Chang AY, Walsh DM, Wu X, Sun X, Frosch MP, Selkoe DJ. Enhanced proteolysis of beta-amyloid in APP transgenic mice prevents plaque formation, secondary pathology, and premature death. Neuron 2003; 40: 1087-1093.
- 93. Lemere CA, Beierschmitt A, Iglesias M, Spooner ET, Bloom JK, Leverone JF, Zheng JB, Seabrook TJ, Louard D, Li D, Selkoe DJ, Palmour RM, Ervin FR. Alzheimer's disease Abeta vaccine reduces central nervous system Abeta levels in a non-human primate, the Caribbean vervet. Am J Pathol 2004; 165: 283-297.
- 94. Lemere CA, Maier M, Jiang L, Peng Y, Seabrook TJ. Amyloidbeta immunotherapy for the prevention and treatment of Alzheimer disease: Lessons from mice, monkeys, and humans. Rejuvenation Res 2006; 9: 77-84.
- 95. Li L, Sengupta A, Haque N, Grundke-Iqbal I, Iqbal K. Memantine inhibits and reverses the Alzheimer type abnormal hyperphosphorylation of tau and associated neurodegeneration. FEBS Lett 2004; 566: 261-269.
- 96. Lue LF, Kuo YM, Roher AE, Brachova L, Shen Y, Sue L, Beach T, Kurth JH, Rydel RE, Rogers J. Soluble amyloid beta peptide concentration as a predictor of synaptic change in Alzheimer's disease. Am J Pathol 1999; 155: 853-862.
- 97. Lupo G, Anfuso CD, Assero G, Strosznajder RP, Walski M, Pluta R, Alberghina M. Amyloid beta (1-42) and its beta (25-35) fragment induce in vitro phosphatidylcholine hydrolysis in bovine retina capillary pericytes. Neurosci Lett 2001; 303: 185-188.

- 98. Lyketsos CG, Steinberg M, Tschanz JT, Norton MC, Steffens DC, Breitner JC. Mental and behavioral disturbances in dementia: findings from the Cache Country Study on Memory in Aging. Am J Psychiatry 2000; 157: 708-714.
- 99. Marco S, Skaper SD. Amyloid beta-peptide 1-42 alters tight junction protein distribution and expression in brain microvessels endothelial cells. Neurosci. Lett 2006; 401: 219-224.
- 100. Marr RA, Rockenstein E, Mukherjee A, Kindy MS, Hersh LB, Gage FH, Verma IM, Masliah E. Neprilysin gene transfer reduces human amyloid pathology in transgenic mice. J Neurosci 2003; 23: 1992-1996.
- 101. Martin-Villalba A, Hahne M, Kleber S, Vogel J, Falk W, Schenkel J, Krammer PH. Therapeutic neutralization of CD95-ligand and TNF attenuates brain damage in stroke. Cell Death Differ 2001; 8: 679-686.
- 102. Matsuoka Y, Saito M, LaFrancois J, Saito M, Gaynor K, Olm V, Wangl L, Casey E, Lu Y, Shiratori C, Lemere C, Duff K. Novel therapeutic approach for the treatment of Alzheimer's disease by peripheral administration of agents with an affinity to β -amyloid. J Neurosci 2003; 23: 29-33.
- 103. McCarty MF. Toward prevention of Alzheimer's disease Potential nutraceutical strategies for suppressing the production of amyloid beta peptides. Med Hypotheses 2006; 67: 682-697.
- 104. McLean CA, Cherny RA, Fraser FW, Fuller SJ, Smith MJ, Beyreuther K, Bush AI, Masters CL. Soluble pool of Abeta amyloid as a determinant of severity of neurodegeneration in Alzheimer's disease. Ann Neurol 1999; 46: 860.
- 105. Mizutani T, Sakata M, Enomoto M, Kasahara M, Yamada S.. Pathological heterogeneity of Alzheimer type dementia. In Alzheimer Disease: Biology, Diagnosis and Therapeutics. Iqbal K, Winbland B, Nishimura T, Takeda M, Wisniewski HM (eds.). John Wiley & Sons Ltd., Chichester 1997; pp. 247-255.
- 106. Moore AH, O'Banion MK. Neuroinflammation and anti-inflammatory therapy for Alzheimer's disease. Adv Drug Del Rev 2002; 54: 1627-1656.
- 107. Morihara T, Teter B, Yang F, Lim GP, Boudinot S, Boudinot FD, Frautschy SA, Cole GM. Ibuprofen suppresses interleukin-1beta induction of pro-amyloidogenic alpha1-antichymotrypsin to ameliorate beta-amyloid pathology in Alzheimer's models. Neuropsychopharmacology 2005; 30: 1111-1120.
- 108. Mruthinti S, Buccafusco JJ, Hill WD, Waller JL, Jackson TW, Zamrini EY, Schade RF. Autoimmunity in Alzheimer's disease: increased levels of circulating IgGs binding Abeta and RAGE peptides. Neurobiol Aging 2004; 25: 1023-1032.
- 109. Mudher A, Lovestone S. Alzheimer's disease-do tauists and baptists finally shake hands? Trends Neurosci 2002; 25: 22-26.
- 110. Nawashiro H, Matin D, Hallenbeck JM. Neuroprotective effects of TNF binding protein in focal cerebral ischemia. Brain Res 1997; 778: 265-271.
- 111. Naslund J, Haroutunian V, Mohs R, Davis KL, Davies P, Greengard P, Buxbaum JD. Correlation between elevated levels of amyloid beta-peptide in the brain and cognitive decline. JAMA 2000; 283: 1571-1577.
- 112. Nicoll JA, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO. Neuropathology of human Alzheimer disease after immunization with amyloid-beta peptide: a case report. Nat Med 2003; 9: 448-452.

- 113. Niedermeyer E. Consideration of the ischemic basis and therapy of Alzheimer disease. Clin EEG Neurosci 2007; 38: 55-56.
- 114. Ooboshi H, Ibayashi S, Takada J, Kumai Y, Iida M. Brain ischemia as a potential target of gene therapy. Exp Gerontol 2003; 38: 183-187.
- 115. Oddo S, Billings L, Kesslak JP, Cribbs DH, LaFerla FM. Abeta immunotherapy leads to clearance of early, but not late, hyperphosphorylated tau aggregates via the proteasome. Neuron 2004; 43: 321-332.
- 116. Ohkura T, Isse K, Akazawa K, Hamamoto M, Yaoi Y, Hagino N. Evaluation of estrogen treatment in female patients with dementia of the Alzheimer type. Endocr J 1994; 41: 361-371.
- 117. Orgogozo JM, Gilman S, Dartigues JF, Laurent B, Puel M, Kirby LC, Jouanny P, Dubois B, Eisner L, Flitman S, Michel BF, Boada M, Frank A, Hock C. Subacute meningoencephalitis in a subset of patients with Alzheimer's disease after Abeta42 immunization. Neurology 2003; 61: 46-54.
- 118. Pluta R. Influence of prostacyclin on early morphological changes in the rabbit brain after complete 20-min ischemia. J Neurol Sci 1985; 70: 305-316.
- 119. Pluta R, Kida E, Lossinsky AS, Gołąbek AA, Mossakowski MJ, Wiśniewski HM. Complete cerebral ischemia with short-term survival in rats induced by cardiac arrest. I. Extracellular accumulation of Alzheimer's beta-amyloid protein precursor in the brain. Brain Res 1994; 649: 323-328.
- 120. Pluta R, Barcikowska M, Januszewski S, Misicka A, Lipkowski AW. Evidence of blood-brain barrier permeability/leakage for circulating human Alzheimer's beta-amyloid-(1-42)-peptide. NeuroReport 1996; 7: 1261-1265.
- 121. Pluta R, Misicka A, Januszewski S, Barcikowska M, Lipkowski AW. Transport of human beta-amyloid peptide through the rat blood-brain barrier after global cerebral ischemia. Acta Neurochir (Suppl.) 1997; 70: 247-249.
- 122. Pluta R. Experimental model of neuropathological changes characteristic for Alzheimer's disease. Folia Neuropathol 1997; 35: 94-98.
- 123. Pluta R, Barcikowska M, Misicka A, Januszewski S, Lipkowski AW. Disappearing diffuse amyloid plaques. Neurobiol Aging 1998; 19: S131.
- 124. Pluta R, Barcikowska M, Misicka A, Lipkowski AW, Spisacka S, Januszewski S. Ischemic rats as a model in the study of the neurobiological role of human β -amyloid peptide. Time-dependent disappearing diffuse amyloid plaques in brain. NeuroReport 1999; 10: 3615-3619.
- Pluta R. The role of apolipoprotein E in the deposition of betaamyloid peptide during ischemia-reperfusion brain injury. A model of early Alzheimer's disease. Ann NY Acad Sci 2000; 903: 324-334.
- 126. Pluta R, Misicka A, Barcikowska M, Spisacka S, Lipkowski AW, Januszewski S. Possible reverse transport of beta-amyloid peptide across the blood-brain barrier. Acta Neurochir (Suppl) 2000; 76: 73-77.
- 127. Pluta R. Blood-brain barrier dysfunction and amyloid precursor protein accumulation in microvascular compartment following ischemia-reperfusion brain injury with 1-year survival. Acta Neurochir (Suppl) 2003; 86: 117-122.

- 128. Pluta R. Alzheimer lesions after ischemia-reperfusion brain injury. Folia Neuropathol 2004; 42: 181-186.
- 129. Pluta R. From brain ischemia-reperfusion injury to possible sporadic Alzheimer's disease. Curr Neurovasc Res 2004; 1: 441-453.
- 130. Pluta R. Pathological opening of the blood-brain barrier to horseradish peroxidase and amyloid precursor protein following ischemia-reperfusion brain injury. Chemotherapy 2005; 51: 223-226.
- 131. Pluta R, Ułamek M, Januszewski S. Micro-blood-brain barrier openings and cytotoxic fragments of amyloid precursor protein accumulation in white matter after ischemic brain injury in long-lived rats. Acta Neurochir (Suppl) 2006; 96: 267-271.
- 132. Pluta R. Ischemia-reperfusion factors in sporadic Alzheimer's disease. In: New Research on Alzheimer's disease. Welsh EM (ed.). Nova Science Publishers, New York 2006; pp.183-234.
- 133. Pluta R. Is the ischemic blood-brain barrier insufficiency responsible for full-blown Alzheimer's disease? Neurol Res 2006; 28: 266-271.
- 134. Pluta R, Ułamek M. Brain amyloidosis following ischemiareperfusion injury. Curr Trends Neurol 2006; 2: 41-46.
- 135. Pluta R. Role of ischemic blood-brain barrier on amyloid plaques development in Alzheimer's disease brain. Curr Neurovasc Res 2007; 4: 121-129.
- 136. Pluta R. Is the ischemic blood-brain barrier a Trojan horse in Alzheimer's disease brain? In: Ischemia-reperfusion pathways in Alzheimer's disease. Pluta R (ed.). Nova Science Publishers, Inc., New York 2007; pp.139-184.
- 137. Pluta R. Ischemia-reperfusion pathways in Alzheimer's disease. Nova Science Publishers, Inc., New York 2007.
- Pluta R, Ułamek M. New proposals for treatment sporadic Alzheimer's disease. Cent Nerv Syst Agents Med Chem 2008; 8: 286-296.
- 139. Pluta R, Januszewski S, Ułamek M. Ischemic blood-brain barrier and amyloid in white matter as etiological factors in leukoaraiosis. Acta Neurochir Suppl 2008; 102: 353-356.
- 140. Pluta R, Ułamek M. Brain ischemia and ischemic blood-brain barrier as etiological factors in sporadic Alzheimer's disease. Neuropsychiatr Dis Treat 2008; 4: 855-864.
- 141. Pluta R, Ułamek M, Jabłoński M. Alzheimer's mechanisms in ischemic brain degeneration. Anat Rec 2009; 292: 1863-1881.
- 142. Pluta R, Januszewski S, Jabłoński M, Ułamek M. Factors in creepy delayed neuronal death in hippocampus following brain ischemia-reperfusion injury with long-term survival. Acta Neurochir Suppl 2010; 106: 37-41.
- 143. Pogue Al, Lukiw WJ. Angiogenic signaling in Alzheimer's disease. NeuroReport 2004; 15: 1507-1510.
- 144. Price DL, Sisodia SS. Mutant genes in familial Alzheimer's disease and transgenic models. Annu Rev Neurosci 1998; 21: 479-505.
- 145. Rakover I, Arbel M, Solomon B. Immunotherapy against APP β -secretase cleavage site improves cognitive function and reduces neuroinflammation in Tg2576 mice without a significant effect on brain A β levels. Neurodegener Dis 2007; 4: 392-402.

- 146. Ramirez BG, Blazquez C, Gomez del Pulgar T, Guzmain M, de Ceballos ML. Prevention of Alzheimer's disease pathology by cannabinoids: neuroprotection mediated by blockade of microglial activation. J Neurosci 2005; 25: 1904-1913.
- 147. Rangan SK, Liu R, Brune D, Planque S, Paul S, Sierks MR. Degradation of beta-amyloid by proteolytic antibody light chains. Biochemistry 2003; 42: 14328-14334.
- 148. Reisberg B, Doody R, Stoffler A, Schmitt F, Ferris S, Mobius HJ, and Memantine Study Group. Memantine in moderate-tosevere Alzheimer's disease. N Engl J Med 2003; 348: 1333-1341.
- 149. Roberts SB. Gamma-secretase inhibitors and Alzheimer's disease. Adv Drug Deliv Rev 2002; 54: 1579-1588.
- 150. Rosenblum WI, Murata S, Nelson GH, Werner PK, Ranken R, Harmon RC. Anti-CD31 delays platelet adhesion/aggregation at sites of endothelial injury in mouse cerebral arterioles. Am J Pathol 1994; 145: 33-36.
- 151. Saito K, Suyama K, Nishida K, Sei Y, Basile AS. Early increases in TNF-alpha, IL-6 and IL-1 beta levels following transient cerebral ischemia in gerbil brain. Neurosci Lett 1996; 206: 149-152.
- 152. Sastre M, Dewachter I, Rossner S, Bogdanovic N, Rosen E, Borghgraef P, Evert BO, Dumitrescu-Ozimek L, Thal DR, Landreth G, Walter J, Klockgether T, Van Leuven F, Heneka MT. Nonsteroidal anti-inflammatory drugs repress beta-secretase gene promotor activity by the activation of PPARgamma. Proc Natl Acad Sci USA 2006; 103: 443-448.
- 153. Saver JL. Time is brain quantified. Stroke 2006; 37: 263-266.
- 154. Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Liao Z, Lieberburg I, Motter R, Mutter L, Soriano F, Shopp G, Vasquez N, Vandevert C, Walker S, Wogulis M, Yednock T, Games D, Seubert P. Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. Nature 1999; 400: 173-177.
- 155. Schmitt FA, Davis DG, Wekstein DR, Smith CD, Ashford JW, Markesbery WR. "Preclinical" Alzheimer's disease revisited: neuropathology of cognitively normal older adults. Neurology 2000; 55: 370-376.
- 156. Schott JM, Price SL, Frost C, Whitwell JL, Rossor MN, Fox NC. Measuring atrophy in Alzheimer disease – a serial MRI study over 6 and 12 months. Neurology 2005; 65: 119-124.
- 157. Schumock GT. Economic considerations in the treatment and management of Alzheimer's disease. Am J Health Syst Pharm 1998; 55 Suppl 2: S17-S21.
- 158. Selkoe DJ. Clearing the brain's amyloid cobwebs. Neuron 2001; 32: 177-180.
- 159. Selkoe DJ. Alzheimer's disease results from the cerebral accumulation and cytotoxicity of amyloid beta-protein. J Alzheimers Dis 2001; 3: 75-80.
- 160. Seubert P, Vigo-Pelfrey C, Esch F, Lee M, Dovey H, Davis D, Sinhas S, Schlossmacher M, Swindlehurst C, McCormack R, Wolfertt R, Selkoe D, Liberburg I, Schenk D. Isolation and quantification of soluble Alzheimer's beta-peptide from biological fluids. Nature 1992; 359: 325-327.
- 161. Snowdon DA, Greiner LH, Mortimer JA, Riley KP, Greiner PA, Markesbery WR. Brain infarction and the clinical expression of Alzheimer disease. The Nun Study JAMA 1997; 277: 813-817.

- 162. Snowdon DA. Healthy aging and dementia: findings from the nun study. Ann Int Med 2003; 139: 450-454.,
- 163. Sohrabji F. Guarding the blood-brain barrier: A role for estrogen in the etiology of neurodegenerative disease. Gene Expression 2007; 13: 311-319.
- 164. Soto C. Protein misfolding and disease; protein refolding and therapy. FEBS Lett 2001; 498: 204-207.
- 165. Spencer B, Rockenstein E, Crews L, Marr R, Masliah E. Novel strategies for Alzheimer's disease treatment. Expert Opin Biol Ther 2007; 7: 1853-1867.
- 166. Spires TL, Hyman BT. Transgenic models of Alzheimer's disease: learning from animals. NeuroRx 2005; 2: 423-437.
- 167. Spisacka S, Pluta R. Demographic and epidemiological profile of patients with Alzheimer's disease. Ann Univ M Curie-Skłodowska (Suppl. XIII) 2003; LVIII: 157-162.
- 168. Swerdlow RH, Khan SM. A "mitochondria cascade hypothesis" for sporadic Alzheimer's disease. Med Hypotheses 2004; 63: 8-20.
- 169. Swerdlow RH. Is aging part of Alzheimer's disease, or is Alzheimer's disease part of aging. Neurobiol Aging 2007; 28: 1465-1480.
- 170. Szekely CA, Thome JE, Zandi PP, Messias E, Breitner JC, Goodman SN. Nonsteroidal anti-inflammatory drugs for the prevention of Alzheimer's disease: a systematic review. Neuroepidemiology 2004; 23: 159-169.
- 171. Takada J, Ooboshi H, Ago T, Kitazono T, Yao H, Kadomatsu K, Muramatsu T, Ibayashi S, Iida M. Postischemic gene transfer of midkine, a neurotrophic factor, protects against focal brain ischemia. Gene Ther 2005; 12: 487-493.
- 172. Tanzi RE, Moir RD, Wagner SL. Clearance of Alzheimer's Abeta peptide: the many roads to perdition. Neuron 2004; 43: 605-608.
- 173. Tariot PN, Farlow MR, Grossberg GT, Graham SM, McDonald S, Gergel I, and Memantine Study Group. Memantine treatment in patients with moderate to severe Alzheimer disease already receiving donepezil: a randomized controlled trail. JAMA 2004; 291: 317-324.
- 174. Tarkowski E, Issa R, Sjogren M, Wallin B, Blennow K, Tarkowski A, Kumar P. Increased intrathecal levels of the angiogenic factors VEGF and TGF-beta in Alzheimer's disease and vascular dementia. Neurobiol Aging 2002; 23: 237-243.
- 175. Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, Hill R, Hansen LA, Katzman R. Physical basis of cognitive alterations in Alzheimer's disease: synapse loss is the major correlate of cognitive impairment. Ann Neurol 1991; 30: 572-580.
- 176. Thomas T, Thomas G, McLendon C, Sutton T, Mullan M. beta-Amyloid-mediated vasoactivity and vascular endothelial damage. Nature 1996; 380: 168-171.
- 177. Vermeer SE, Prins ND, den Heijer T, Hofman A, Koudstaal PJ, Breteler MMB. Silent brain infarcts and the risk of dementia and cognitive decline. N Engl J Med 2003; 348: 1215-1222.
- 178. Vukovic V, Lovrencic-Huzjan A, Solter VV, Dordevic V, Demarin V. Hormone replacement therapy is there a place for its use in neurology? Coll Antropol 2003; 27: 413-424.
- 179. Waring SC, Rocca WA, Petersen RC, O'Brien PC, Tangalos EG, Kokmen E. Postmenopausal estrogen replacement therapy and risk of AD: a population-based study. Neurology 1999; 52: 965-970.

- 180. Wen Y, Yang SH, Liu R, Perez EJ, Brun-Ziukemagel AM, Koulen P, Simpkins JW. Cdk5 is involved in NFT-like tauopathy induced by transient cerebral ischemia in female rats. Biochim Biophys Acta 2007; 1772: 473-483.
- 181. Willing AE, Cuevas J, Pennypacker KR. Treatment of Alzheimer's disease: New insights from treatment of stroke at delayed time points. In: Ischemia-reperfusion pathways in Alzheimer's disease. Pluta R (ed.). Nova Science Publishers, Inc., New York 2007; pp. 185-203.
- Wiśniewski HM, Maslińka D. Beta-protein immunoreactivity in the human brain after cardiac arrest. Folia Neuropathol 1996; 34: 65-71.
- 183. Xia CF, Yin H, Borlongan CV, Chao L, Chao J. Kallikrein gene transfer protects against ischemic stroke by promoting glial cell migration and inhibiting apoptosis. Hypertension 2004; 43: 452-459.
- 184. Xie CW. Calcium-regulated signaling pathways: role in amyloid beta-induced synaptic dysfunction. Neuromolec Med 2004; 6: 53-64.
- 185. Xuereb JH, Brayne C, Dufouil C, Gertz H, Wischik C, Harrington C, Mukaetova-Ladinska E, Mc Gee MA, O'Sullivan A, O'Connor D, Paykel ES, Huppert FA. Neuropathological findings in the very old. Results from the first 101 brains of a population-based longitudinal study of dementing disorders. Ann NY Acad Sci 2000; 903: 490-496.
- 186. Yang F, Lim GP, Begum AN, Ubeda OJ, Simmons MR, Ambegaokar SS, Chen PP, Kayed R, Glabe CG, Frautschy SA, Cole GM. Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces amyloid in vivo. J Biol Chem 2005; 280: 5892-5901.
- 187. Yang SH, Simpkins JW. Ischemia-reperfusion promotes tau and beta-amyloid pathology and a progressive cognitive impairment. In: Ischemia-reperfusion pathways in Alzheimer's disease. Pluta R (ed.). Nova Science Publishers, Inc., New York 2007; pp.113-138.
- 188. Youdim MB, Buccafusco JJ. Multi-functional drugs for various CNS targets in the treatment of neurodegenerative disorders. Trends Pharmacol Sci 2005; 26: 27-35.
- 189. Zhang RL, Chopp M, Liu Y, Zaloga C, Jiang N, Jones ML, Miyasaka M, Ward PA. Anti-ICAM-1 antibody reduces ischemic cell damage after transient middle cerebral artery occlusion in the rat. Neurology 1994; 44: 1747-1751.
- 190. Zhao H, Yenari MA, Cheng D, Barreto-Chang OL, Sapolsky RM, Steinberg GK. Bcl-2 transfection via herpes simplex virus blocks apoptosis-inducing factor translocation after focal ischemia in the rat. J Cereb Blood Flow Metab 2004; 24: 681-692.
- 191. Zipser BD, Johanson CE, Gonzalez L, Berzin TM, Tavares R, Hultte CM, Vitek MP, Hovanesian V, Stopa EG. Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. Neurobiol Aging 2007; 27: 977-986.
- 192. Zlokovic BV, Martel CL, Matsubara E, McComb JG, Zhang G, McCluskey RT, Frangione B, Ghiso J. Glycoprotein 330/megalin: probable role in receptor-mediated transport of apolipoprotein J alone and in a complex with Alzheimer disease amyloid beta at the blood-brain and blood-cerebrospinal fluid barriers. Proc Natl Acad Sci USA 1996; 93: 4229-4234.
- 193. Zlokovic BV. Neurovascular mechanisms of Alzheimer's neurodegeneration. Trends Neurosci 2005; 28: 202-208.